REMARKS

In the Final Action dated July 10, 2009, Claims 1 and 3-29 were pending in the application. Claims 13-27 were withdrawn from further consideration as drawn to non-elected inventions. Claims 1 and 3-12 were under examination and are rejected. Specifically, claims 1, 3 and 7-9 were rejected under 35 U.S.C. §102(b) as anticipated by Maliszewski (*Pathol. Biol.* 2001; 49: 481-483). Claims 1 and 3-12 were rejected under 35 U.S.C. §112, first paragraph, as failing to comply with the enablement requirement.

This Response addresses each of the Examiner's rejections. Applicants therefore respectfully submit that the present application is in condition for allowance. Favorable consideration of all pending claims is therefore respectfully requested.

Claim Amendments

By way of the foregoing amendments, Applicants have amended claim 1 to recite "delaying onset", instead of "preventing onset". Support for this amendment is found in the specification, e.g., page 1, line 9. Claim 1 has also been amended to define the autoimmune disease to be specially "diabetes", a feature previously delineated in dependent claim 12. Finally, claim 1 has been amended to add "which delays onset of diabetes" at the end of the claim to refer back to the preamble. No new matter is introduced.

Claim 4 has been canceled, without prejudice.

Claims 11-12 have been canceled, without prejudice, in light of the amendments made to claim 1.

Applicants reserve the right to pursue the subject matter as originally claimed in a continuation application.

35 U.S.C. §102(b)

Claims 1, 3 and 7-9 are rejected under 35 U.S.C. §102(b) as anticipated by Maliszewski (Pathol. Biol. 2001; 49: 481-483).

Applicants first observe that claims 4-6 and 10-12 were not included in the rejection.

Because independent claim 1 has been amended, *inter alia*, to incorporate the feature of claim 12, Applicants respectfully submit that the anticipation rejection is obviated in light of the amendments to claim 1.

Further, Applicants respectfully submit that Maliszewski does not teach, in any event, administration of Flt-3L in an amount effective to increase a sub-type of non-activated, immature and tolerogenic DC selected from Plasmacytoid DC, CD8⁺ DC or their equivalents, thereby inducing or maintaining immune tolerance in the subject which delays onset of diabetes, as presently claimed.

Accordingly, the rejection under 35 U.S.C. §102(b) based on Maliszewski is overcome. Withdrawal of the rejection is therefore respectfully requested.

35 U.S.C. §112, First Paragraph

Claims 1 and 3-12 are rejected under 35 U.S.C. §112, first paragraph, as allegedly failing to comply with the enablement requirement.

Applicants wish to draw the Examiner's attention to the fact that the claims, as amended, are directed to methods of delaying onset of diabetes. Applicants respectfully submit that the claims, as presently recited, are fully enabled by the specification. Applicants will address the issues raised by the Examiner as follows.

The Examiner states that the data provided in the specification (e.g., Example 12) were based on a mouse model of type 1 diabetes, and would not apply to any other autoimmune

diseases. This aspect of the rejection is obviated in light of the amendment to claim 1 to define the disease as diabetes.

Further, the Examiner contends that Applicants' conclusion that repeated administration of mFL prevents diabetes is solely based on the observation of elevated blood sugar, which, according to the Examiner, does not correlate to an equivalent destruction of islet cells. Moreover, referring to page 44, lines 5-16 of the specification (Example 12), the Examiner notes that the experiment therein used NOD mice and demonstrates numerous different times of administrating murine Flt3L which did not *prevent* diabetes. Therefore, the Examiner is of the opinion that while the specification seems to support *delaying* some of the *complications* related to type 1 diabetes (e.g. hyperglycemia) through administration of Flt3L, this is not the same as *preventing onset of the disease itself*.

In the first instance, Applicants respectfully submit that the claims have been amended to recite "delaying" the onset of diabetes, instead of "preventing". The data exemplified in both Examples 11 and 12, including the data on page 44, lines 5-16 referenced by the Examiner, fully support delaying the onset of diabetes based on administration of mFL.

Furthermore, with respect to the Examiner's position on hyperglycemia vis-a-vis diabetes, Applicants respectfully submit that the NOD mouse is a well-established and accepted model of Type 1 diabetes. It is also well established that the hyperglycemia that develops is a direct and inevitable consequence of an autoimmune attack which destroys the insulin producing cells of the islets of the pancreas. See Lampeter et al., Diabetologia 32: 703-708 (1989) (provided hereto as Exhibit 1); especially page 703, column 1, 2nd sentence: "Diabetes develops ... with an onset characterized by ... hyperglycemia ..."). Thus, Applicants respectfully submit that it is well accepted in the art and perfectly valid to use hyperglycemia as the primary measure

to detect diabetes in NOD mice. Moreover, the present application discloses that when mice are treated with Flt3L, this treatment can protect the mice, i.e., the lack of hyperglycemia is correlated with the lack of autoimmune destruction of the pancreatic islets. This was demonstrated by histology. For example, the specification on page 43 (Example 11) shows that the mice classed as diabetic on the basis of being hyperglycemic had only 9 islets per 7 pancreatic sections, as opposed to 35 in the mice that had not yet become hyperglycemic. The mice that were found to be protected from becoming hyperglycemic by Flt3L administration had 47 islets per 7 sections, so their islets had not been destroyed.

The Examiner has also made certain comments in relation to insulitis (mononuclear infiltration) on page 11, lines 8-14 of the Action. The Examiner appears to correlate insulitis with diabetes, and concludes that the data provided in the specification (page 47, lines 28-29) would be evidence that the disease had not been prevented.

Applicants respectfully submit that the Examiner's understanding in this regard is incorrect. Applicants direct the Examiner's attention to Lampeter et al., page 705, column 1, 2nd to last sentence of paragraph 1, which confirms that diabetes in NOD mice has a strong female preponderance, but insulitis is present to a similar degree in both sexes, i.e., in male mice without diabetes. It is also stated in Lampeter et al. (last paragraph on page 707) that insulitis is in progress well before overt hyperglycemia and complete beta cell destruction. Consistently, in the study disclosed in the present application, the NOR control mice, which were chosen as a closely matched mouse strain which do not develop diabetes or hyperglycemia, nevertheless had mononuclear infiltrates in the pancreas. See page 47, last line, of the specification. Thus, Applicants submit that the fact that insulitis occurred in Flt-3L treated mice (and in control mice) does not negate the protective effects of Flt-3L in delaying the onset of diabetes.

Finally, the Examiner has also commented on co-administration of Flt-3L and a Tolllike receptor ligand. In an effort to advance prosecution, Applicants have canceled claim 4.

directed to co-administration of Flt-3L and a Toll-like receptor ligand, without prejudice.

In view of the foregoing, Applicants respectfully submit that the present specification

fully enables those skilled in the art to practice the methods, as presently claimed, without undue

experimentation. Thus, reconsideration and withdrawal of the enablement rejection under 35

U.S.C. §112, first paragraph are respectfully requested.

Conclusion

It is firmly believed that the subject application is in condition for allowance, which

action is earnestly solicited.

Respectfully submitted,

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XZ:ab

Enc.: Exhibit 1

EXHIBIT 1



Review

Lessons from the NOD mouse for the pathogenesis and immunotherapy of human Type 1 (insulin-dependent) diabetes mellitus

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Summary. Suitable animal models of human Type 1 (insulindependent) diabetes mellitus have long been sought, in particular a model that would permit detailed histological and immunological investigation of changes in the islet preceding the metabolic disorder. This would allow hypotheses as to pathogenesis of the condition to be examined and interventions such as immunotherapt to be tested. The most widely studied models include the low-dose streptozotocin induced diabetic mouse and the BB rat, but both differ in important respects from the human disease. In this review we describe one highly successful model, the non obese diabetic mouse. Selected aspects of pathogenesis and immunotherapy are presented and analogies with human Type 1 diabetes discussed.

Key words: Non obese diabetic (NOD) mouse, pathogenesis Type 1 (insulin-dependent) diabetes mellitus, immunotherapy Type 1 diabetes.

The NOD mouse was derived from a cataract-developing substrain of the outbred JcI-ICR mouse by selective breeding from 1974 to 1980 [1]. Diabetes develops spontaneously between the 12th and 30th week of age, with an onset characterized by polydipsia, glycosuria, rapid weight loss, hyperglycaemia and ketoacidosis (Table 1). The onset of hyperglycaemia is preceded by insulitis. progressive B-cell destruction and decreasing circulating insulin levels leading to insulin dependency [2-4]. Without insulin treatment the animals die within 4 to 8 weeks (unpublished observations). Thus, clinical and pathological features in the NOD mouse closely resemble human Type 1 (insulin-dependent) diabetes mellitus. Since all conclusions drawn from animal models are, however, based on analogy with human disease, the analogy needs detailed validation. For this reason we describe similarities and differences relating to pathogenesis and immunotherapy in the NOD mouse and human Type 1 diabetes.

Genetic background

Continued in-breeding of the strain has resulted in high genetic uniformity as shown by morphology, allele distribution of enzymes and other proteins, and immunological studies including mixed lymphocyte reaction and skin grafting [5]. Based on this, the genetic background of insultits and overt diabetes has been investigated by backcross experiments with C57BL, NZB
mice and a non obese non diabetic subline (NON) of
the same origin as the NOD [6-8]. The results indicate
three recessive diabetogenic genes, two of which are
non MHC-linked. One controls the development of
severe insultis and appears to be incompletely dominant, and the other is involved in the progression to
diabetes, probably mediated by a lack of specific suppressor cells. The third, MHC linked, gene is not required for insultits but apparently influences the autoimmune response [7]. It has been suggested that the
NOD mouse has a unique class II MHC, which may
lead to the autoimmune insultits [9]. Furthermore, treat-

Table 1. Comparison of clinical features at onset of diabetes in the human and the NOD mouse

	Type 1 (insulin-dependent) diabetes mellitus		
	human	NOD mouse	
Weight loss	Present	Present	
Polydipsia	Present	Present	
Polyuria	Present	Present	
Hyperglycaemia	> 15 mmol/l	20-30 mmol/1	
Ketoacidosis	Common	Less severe	
Serum insulin	Very low	Very low	
Outcome without insulin	Lethal	Lethal	
Sex preponderance	Female ≥ male	Female > male	

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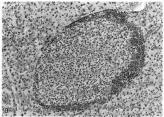




Fig. 1a and b. Micrographs of islets of Langerhans in a 20-week-old female NOD-mouse showing periinsulitis (a), followed by invasion of the islet by lymphocytes penetrating the capsule (b) (Magnification 145 ×. haematoxviin and eosin staining)

ment with anti-I-A monoclonal antibody prevented diabetes in NOD mice [10].

Human Type 1 diabetes is associated with MHC products [11], most closely linked with the HLA DQ region [12]. In mice the equivalent to DQ-beta is the A-β chain, and this has interesting similarities with human diabetes [13]. Back-cross experiments have shown that homozygosity at this gene is necessary for the development of diabetes. The NOD A-β allele is unique in the species in having serine in position 57 instead of aspartic acid (Asp). Similarly in humans DQ-β Asp 57 negative homozygosity is found in 90% of Caucasian Type 1 diabetic patients, whereas, Asp 57 positive homozygosity at DQ-β gives almost complete protection from Type 1 diabetics [13].

Despite strong evidence for an association with a genetic factor or factors, the concordance rate for Type 1 diabetes is surprisingly low in identical twins (30-50%), suggesting that susceptibility is inherited rather than the expressed disease [14]. The NOD mouse resembles man in this respect, since the animals are genetically identical but not all develop diabetes. Abnormal immunological parameters including islet cell antibodies (ICA) and increased numbers of circulating activated T cells are, however, concordant in human twin studies [15, 16] and NOD mice also have a concordant immunological process, as shown by the fact that all females and more than 90% of males exhibit insulitis [7, 17, 18]. The incidence of diabetes is, however, at least twice as high in female NOD mice than in males, and castration experiments suggest that this difference is related to female sex hormones [17]. Castration of mice up to the age of 7 weeks results in an increase of diabetes incidence in males and a decrease of incidence in females.

Administration of testosterone prevents diabetes in castrated animals, whereas oestradiol raises the incidence of diabetes in castrated animals of both sexes. The rate of development of diabetes is also influenced by diet [17]. Although hormonal and dietary influences have not been shown in human diabetes, human Type 1 diabetes and the NOD model suggest that genetic factors predispose to the autoimmune disorder, but have limited importance for clinical expression of the disease.

The genetic uniformity in this inbred strain has great advantages in the experimental situation as a guarantee of identity, but for the same reason has limited relevance to the human situation. Thus, even though 95% of Type 1 diabetic patients are HLA DR3 or DR4 positive [20], there is marked genetic heterogeneity. The mechanism of inheritance of the disease in the NOD mouse cannot, therefore, be identical to that in man, although it constitutes one of the possible alternatives.

Histopathology

Insulitis is the pathological hallmark recent onset Type 1 diabetes and is observed in the NOD mouse from at least the 4th week of age [21, 22]. The earliest change is periinsulitis adjacent to the pancreatic ducts, followed by invasion of the islet capsule by small lymphocytes which penetrate the islet (Fig. 1). The final stage is characterized by small islets from which B-cells have disappeared, with resolution of insulitis. The different stages of this process can, however, be found within the same pancreas at any age. Phenotyping of lymphocyte subsets involved in the insulitis has produced conflicting results [23-26]. We have found that monocytes and B-lymphocytes are the predominant cell population [25]. Previous studies have reported L3T4 cells (mainly helper/inducer) and MHC class-II cells as the most represented subsets [23, 26]. Within the T-lymphocyte population L3T4 cells are more frequently found than Lyt-2 cells (mainly cytotoxic/suppressor) [24].

Table 2. Comparison of morphological features in human diabetes and the NOD mouse

	Type 1 (insulin-dependent) diabetes mellitus	
	Human	NOD mouse
Periinsulitis/insulitis	Present	Present
Insulitis in subjects without diabetes	?	Present
Small islets lacking B-cells at the end stages	Present	Present
Lymphocytic infiltration in other organs	Rare	Present

The prevalence of insulitis is high in humans who have died soon after the onset of Type 1 diabetes, and in one early study insulitis was present in 16 of 23 who died within 6 months of onset [27, 28] and in 47 out of 60 patients with a diabetes duration of less than one year [29]. The smouldering nature of the process is equally apparent, with normal islets, insulitis and "endstage" islets depleted of B cells within the same histological field. While there is still some controversy concerning the prevalence of insulitis in man, there is agreement on the histological pattern of lymphocytic infiltration (Table 2). As in the NOD mouse, insulitis develops in man as periinsulitis and progresses to infiltration of the islets and B-cell destruction [28]. There has been only one report concerning the phenotype of lymphocyte subsets, based on the pancreas of a child who died at the time of diagnosis [30]. The majority of infiltrating lymphocytes were T-cells, predominantly CD8 positive although other inflammatory cells were present. Thus, despite possible differences between the lymphocyte subsets infiltrating the islets, the NOD mouse is a good model from the histopathological point of view. Diabetes has a strong female preponderance in the NOD mouse (70% vs 20% at 30 weeks of age) but insulitis is present to a similar degree in both sexes. Thus, about 80% of males and 30% of females show insulitis without developing diabetes up to the 30th week of age [4]. In the human situation we remain ignorant as to the time of development of insulitis prior to the disease, although it is assumed to coincide with the appearance of ICA and other autoimmune markers, and it is not known whether individuals with insulitis inevitably progress to diabetes.

In NOD mice lymphocytic infiltration is not restricted to the islets but occurs also in salivary tissue [31] and occasionally in the thyroid and adrenal glands [32], suggesting a wider disturbance of immune tolerance in this animal. Type 1 diabetes is also associated with overt polyendocrine disease and there is an increased prevalence of autoantibodies to thyroid, adrenal or gastric parietal cells, although figures concerning this vary [33]. Infiltration of salivary glands has not, however, been described in human diabetes.

Immunological observations

Autoantibodies

ICA have been found in about 50% of NOD mice up to the 21st week of age but tend to disappear later [34, 35]. Islet cell surface antibodies (ICSA) appear at 3-6 weeks, reach peak incidence and titre at around 12-18 weeks, and decline thereafter [26, 34]. There is no evidence that these autoantibodies are directly involved in B-cell destruction, and both ICA and ICSA might be secondary to islet cell destruction and massive release of cellular antigen, a view which accords with the time course of insulitis in this animal model. Insulin autoantibodies (IAA) have also been reported; they may antedate insulitis [35] and are present in almost all animals later in life [34]. Finally, autoantibodies which immunoprecipitate a 64.000 mol.wt.islet antigen have recently been described [36]. As in humans, the pathogenetic relevance of these autoantibodies remains uncertain, and the prognostic significance of ICA, ICSA, insulin and 64 kilodalton autoantibodies has vet to be investigated in the NOD mouse.

Cell mediated immunity

Several successful attempts have been made to transfer insultis and diabetes via lymphocytes derived from NOD mice, using a variety of protocols. Diabetes appears within a few weeks of lymphocyte transfusion, providing further support for the autoimmune hypothesis. Recipient animals were either newborn or very young normal NOD mice 137, totally irradiated NOD mice 138, 39, or athymic nude mice of NOD origin [40]. Despite these differences, similar results were obtained with regard to the age of lymphocyte donors, and 100% successful transfer of diabetes/insultits can only be achieved with lymphocytes from mice at least 16-19- weeks-old [37, 39]. Interestingly the transfer can be made with either diabetic or non-diabetic donor lym

Table 3. Comparison of immunological features of Type 1 (insulindependent) diabetes in humans and the NOD mouse

	Type 1 diabetes mellitus	
	Human	NOD mouse
Insulin autoantibodies	Present	Present
Islet cell antibodies	Present	Present
Islet cell surface antibodies	Present	Present
Islet cell specific cellular immunity	Present	Present
Abnormal T-helper/T-suppressor	Present	Present
Major T-lymphocyte subset in the insulitis	CD 8+	L3T4+
Aberrant expression of class II MHC on insulin positive cells	Present	?

phocytes. In older non-diabetic recipients (i.e. > 25 weeks) the transfer is much less effective and only 5 of 16 developed diabetes [39]. When separated lymphocyte subsets were used, both Lyt-2* (mainly cytotoxic/suppressor) and L3T4* (mainly helper/inducer) cells appeared to be necessary. In addition, both subsets should be derived from a donor of appropriate age (16 to 19 weeks) as shown in transfer experiments in which L3T4* cells from an appropriate donor over reconstituted with Lyt 2*cells from a 6-week-old donor (or vice versa) but failed to induce diabetes when transferred. It was further shown that newborn mice are susceptible to transfer until the 3rd week (females) and the 5th week (males).

The NOD mouse, therefore, appears susceptible to the transfer of diabetes until the time at which insulitis develops spontaneously. At approximately 16 to 19 weeks the animals acquire the ability to transfer the disease with lymphocytes, but this capacity is often lost in non-diabetic mice from the 25th week onwards. These findings may reflect time dependent differences in the development of necessary lymphocyte subsets (i.e. Thelper/inducer first, antigen-specific effector second and T-suppressor cells, last). If this is the case B-cells might disappear too rapidly for the induction of T-suppressor cells in animals which develop diabetes, whereas animals with slower destruction of B cells may produce sufficient specific T-suppressor cell activity to protect themselves from further B-cell loss.

A variety of cellular cytotoxicity systems have been investigated in search of an active effector cell mechanism in the NOD mouse. Direct cellular cytotoxicity (CTL) was increased as compared to ICR mice using Balb/c islets as targets in a chromium release assay. Antibody dependent cellular cytotoxicity (ADCC) and natural killer (NK) cell activity have been tested in NOD and ICR mice. Both ADCC against chicken erythrocytes in the presence of anti-chicken erythrocyte antibodies and NK activity against Chang liver cells are decreased in the NOD mouse [41]. Another interesting observation is that athymic nude mice with NOD background [40] or NOD mice undergoing neonatal thymectomy [18] did not develop insulitis and diabetes - suggesting a pivotal role for T-lymphocytes in the autoimmune process. Administration of monoclonal antibodies (mAb) specific to some lymphocyte surface markers can block function or destroy the corresponding cell subset. Thus, treatment with anti Thy 1.2 mAb (T cells) prevents diabetes but does not influence the progression of insulitis [40]. Administration of L3T4 mAb abolishes insulitis and diabetes [42, 43]. In addition. Lyt 2+cells (suppressor/cytotoxic) and macrophages are necessary for the development of insulitis since treatment with anti-Lyt2 antibody and silica particles prevents B-cell destruction [44].

Thus, macrophages and Lyt2⁺ cells are required for induction of the autoimmune process by appropriate antigen presentation and for generation of specific ef-

fector cells, respectively. On the other hand, cyclophosphamide (known to impair T-suppressor cells) promotes overt diabetes and increases its incidence in the NOD mouse [45]. These data suggest the presence of specific T-suppressor cells in the NOD mouse, although these are clearly not efficient enough to maintain tolerance in all cases.

Aberrant expression of HLA class II on B cells has been claimed to play an important role in the initiation of the autoimmune process leading to diabetes [46]. Class II expression was found in a child who died soon after clinical presentation [30] and confirmed by an immunohistological study of formalin fixed paraffin embedded tissue from post mortem cases with recent onset of diabetes [47]. In the NOD mouse conflicting results have been obtained. Hanafusa et al. [48] described aberrant expression of class II molecules prior to insulitis, as identified by anti-IA mouse antibodies, not only in the NOD but also to some extent in BALB/C and B10.GD mice. These results could not be confirmed using P7/7 rat MAb [23] which recognizes class-II molecules of b, d and k haplotype [49], and all islet cells appear negative with this antibody [25].

Immunotherapy

Cyclosporin A reduces insulitis in the NOD mouse but is unable to abolish it [50], while ICSA titres were similar or even higher than in control animals. In another reported against insulitis [51]. These data indicate that cyclosporin A can partially suppress the cell mediated reaction but not the production of ICSA. Unfortunately the incidence of diabetes in cylcosporin treated animals was not investigated.

Nicotinamide, an inhibitor of poly-ADP-ribose synthetase, reduces the incidence of insulitis and diabetes in the NOD mouse [52]. Cyclophosphamide increases the incidence of diabetes but this effect can be blocked by nicotinamide [19]. ADCC is naturally elevated in the NOD mouse but falls after nicotinamide treatment [53]. This suggests an important role for ADCC and also indicates that nicotinamide has immunomodulatory properties. This is supported by the observation that single injections of nicotinamide prior to allogeneic islet transplantation prolong graft survival, a treatment which was much more successful if nicotinamide was combined with desferroxamine, an iron-chelating agent [54]. Nicotinamide seems to have some benefit in newly diagnosed Type 1 diabetic patients [55, 56] and increases C-peptide secretion in the first year after diagnosis [57].

Conclusion

The NOD mouse model shares a number of important characteristics with human Type 1 diabetes. The disease develops spontaneously and is not accompanied by general immunodeficiency as in the BB rat. Differences include simultaneous lymphocytic infiltration of salivary glands and other organs, and a strong female predominance. Even so, study of mechanisms involved in insultits, B-cell destruction, and the generation of other immunological disturbances allows hypotheses concerning human Type 1 diabetes to be developed and tested. The availability of high and low incidence lines may, in addition, offer clues to factors involved in the onset of diabetes.

Insultits is in progress well before overt hyperglycaemia in the NOD mouse, and this is important for two reasons: (1) it allows the autoimmune process to be defined before complete B-cell destruction and hyperglycaemia have occurred. This might prove very useful in the search for new markers during this crucial phase of the natural history. (2) the prolonged and well defined prodromal period provides an excellent opportunity to test different approaches to immunotherapy early in the prediabetic stage.

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References

- Tochino Y (1986) Discovery and breeding of the NOD mouse. In: Tarui S, Tochino Y, Nonaka K (eds) Insulitis and type I diabetes

 lessons from the NOD mouse. Academic Press, New York London, pp 3-10
- Ohneda A, Kobayashi T, Nihei J, Tochino Y, Kanaya H, Makino S (1984) Insulin and glucagon in spontaneously diahetic nonobese mice. Diabetologia 27: 460-463
- Kolb H (1987) Mouse models of insulin dependent diabetes: low-dose streptozotocin-induced diabetes and nonobese diabetic (NOD) mice. Diab Metab Rev 3: 751-778
- Tochino T (1987) The NOD mouse as a model of Type 1 diabetes. CRC Crit Rev Immunol 8: 49-81
- Komeda K, Goto N (1986) Genetic monitoring of the NOD mouse. In: Tarui S, Tochino Y, Nonaka K (eds) Insulitis and type I diabetes - Lessons from the NOD mouse. Academic Press New York London, pp 11-22.
- 6. Makino S, Hayashi Y (1986) Genetic analysis for insulitis in the NOD mouse. In: Tarui S, Tochino Y, Nonaka K (eds) Insulitis and type I diabetes - lessons from the NOD mouse. Academic Press, New York London, pp 23-31
- Wicker LS, Miller BJ, Coker LZ, McNally SE, Scott S, Mullen Y, Appel MC (1987) Genetic control of diabetes and insulitis in the nonobese diabetic (NOD) mouse. J Exp Med 165: 1639–1654
- Prochazka M, Leiter EH, Serreze DV, Coleman DL (1987) Three recessive loci required for insulin-dependent diabetes in nonobese diabetic mice. Science 237: 286-289
- Nishimoto H, Kikimoto T, Yamamura KI, Kishimoto T (1987)
 Prevention of autoimmune insulitis by expression of I-B molecules in NOD mice. Nature 328: 432
 444
- Boitard C, Bendelac A, Richard MF, Bach JF (1988) Prevention
 of diabetes in nonobese diabetic mice by anti-I-A monoclonal
 antibodies: transfer of protection by splenic T cells. Proc Natl
 Acad Sci USA 85: 9719-9723
- Nerup J, Platz P, Andersen OO, Christy M, Lyngsoe J, Poulsen JE, Ryder LP, Nielsen LS, Thomsen M, Svejgaard A (1974) HLA antigens and diabetes mellitus. Lancet II: 864-866

- Lernmark Å, Li S, Baekkeskov S, Christie M, Michelsen B, Ursing J, Olsson ML, Sundkvist G (1987) Islet-specific immune mechanisms. Diab Metab Rev 3: 959-980
- Todd JA, Bell JI, McDevitt HO (1987) HLA-DQ beta gene contributes to susceptibility and resistance to insulin-dependent diabetes mellitus. Nature 329: 599-604
- Barnett AH, Eff C, Leslie RDG, Pyke DA (1981) Diabetes in identical twins. Diabetologia 20: 87-93
- Johnston C, Pyke DA, Cudworth AG, Wolf E (1983) HLA-DR typing in identical twins with insulin-dependent diabetes: differences between concordant and discordant pairs. Br Med J 286:
- 253-255

 16. Wolf E, Spencer KM, Cudworth AG (1983) The genetic susceptibility to Type 1 (insulin-dependent) diabetes analysis of the HLA-DR association. Diabetologia 24: 224-230
- Leiter EH, Prochazka M, Coleman DL (1987) Animal model of human disease - the non-obese diabetic mouse. Am J Pathol 128: 380-383
- Ogawa M, Maruyama T, Hasegawa T, Kanaya T, Kobayashi F, Tochino Y, Uda H (1985) The inhibitory effect of neonatal thymectomy on the incidence of insultits in non-obese diabetes (NOD) mice Riomed Res 6: 103-105
- Nakajima H, Tarui S, Tochino Y (1986) Clues to the pathogenesis of diabetes in the NOD mouse based on preventive approaches. In: Tarui S, Tochino Y, Nonaka K (eds) Insultits and type I diabetes - lessons from the NOD mouse. Academic Press, New York London, pp. 181-186
- 20. Maclaren NK (1988) How, when and why to predict IDDM. Diabetes 37: 1591-1594
- Fujita T, Yui R, Kusumoto Y, Serizawa Y, Makino S, Tochino (1982) Lymphocytic insulfits in a non-obese diabetic (NOD) strain of mice: an immunohistochemical and electron microscope investigation. Biomed Res 3: 429-443
- Fujino-Kurihara M, Fujita H, Hakura A, Nonaka T, Tarui S (1985) Morphological aspects on pancreatic islets of non-obese diabetic (NOD) mice. Virchows Arch B 49: 107-120
- Signore A, Cooke A, Pozzilli P, Butcher G, Simpson E, Beverley PCL (1987) Class II and IL2 receptor positive cells in the pancreas of NOD mice. Diabetologia 30: 902-905
- 24. Miyazaki A, Hanafusa T, Yamada K, Miyagawa J, Fujino-Kurihara H, Nakajima H, Nonaka K, Tarui S (1985) Predominance of Tlymphocytes in paucreatic islets and spleen of prediabetic non obese diabetic (NOD) mice: a longitudinal study. Clin Exp Immunol 60: 622-630
- Signore A, Gale EAM, Andreani D, Boverley PCL, Pozzilli P (1989) The natural history of lymphocyte subsets infiltrating the pancreas of NOD mice. Diabetologia 32: 282–289
- Kanazawa Y, Komeda K, Sato S, Mori S, Akanuma K, Takaku F (1984) Non-obese-diabetic mice: immune mechanisms of pancreatic B-cell destruction. Diabctologia 27: 113-115
- Gepts W (1965) Pathologic anatomy of the pancreas in juvenile diabetes mellitus. Diabetes 14: 619-633
- Gepts W (1987) Islet morphologic changes in diabetes. Diab Metab Rev 3: 859-872
- 29. Foulis AK, Liddle CN, Farquharson MA, Richmond JA, Weir RS (1986) The histopathology of the pancreas in Type 1 (insulin-dependent) diabetes mellitus: a 25-year review of death in patients under 20 years of age in the United Kingdom. Diabetologia 29: 267-274
- Bottazzo GF, Dean BM, McNally J, Hackay EH, Swift PGF, Gamble DR (1985) In situ characterization of autoimmune phenomena and expression of HLA molecules in the pancreas in diabetic insulitis. N Engl J Med 313: 353-360
- Miyagawa J, Hanafusa T, Miyazaki A, Yamada K, Fujino-Kurihara H, Nakajima H, Kono N, Nonaka K, Tochino Y, Tariis (1986) Ultrastructural and immunocytochemical aspects of lymphocytic submandibulitis in the non-obese diabetic (NOD) mouse. Virchows Arch B 51: 215-225
- Asamoto H, Oishi M, Akazawa Y, Tochino Y (1986) Histological and immunological changes in the thymus and other organs in NOD mice. In: Tarui S, Tochino Y, Nonaka K (eds) Insulitis and

- type I diabetes lessons from the NOD mouse. Academic Press, New York London, pp 61-71
- Eisenbarth GS, Rassi N (1983) The polyglandulare failure syndromes. In: Davis TF (ed) Autoimmune endocrine disease. John Wiley & Sons, New York, pp 193–206
- Reddy S, Bibby NJ, Elliot RB (1988) Ontogeny of islet cell antibodies, insulin autoantibodies and insulitis in the non-obese diabetic mouse. Diabetologia 31: 322-328
- Pontesilli O, Carotenuto P, Gazda LS, Pratt PF, Prowse SJ (1987) Circulating lymphocyte populations and autoantibodies in nonobese diabetic (NOD) mice: a longitudinal study. Clin Exp Immunol 70: 84-93
- Atkinson MA, MacLaren NK (1988) Autoantibodies in nonobese diabetic mice immunoprecipitate 64,000-M_r islet antigen. Diabetes 37: 1587-1590
- Bendelac A, Carnaud C, Boitard C, Bach JF (1987) Syngenic transfer of autoimmune diabetes from diabetic NOD mice to healthy neonates. J Exp Med 166: 823-832
- Miller BJ, Appel MC, O'Neil JJ, Wicker LS (1988) Both the Lyt-2⁺ and L3T4⁺T cell subset are required for the transfer of disheter in prophere diabetic mice. J Immunol 140: 52.58
- diabetes in nonobese diabetic mice. J Immunol 140: 52-58
 39. Wicker LS, Miller BJ, Mullen Y (1986) Transfer of autoimmune diabetes mellitus with splenocytes from nonobese diabetic (NOD) mice. Diabetes 35: 855-860
- Harada M, Makino S (1986) Immunological manipulation of diabetes production in NOD mice. In: Tarui S, Tochino Y, Nonaka K (eds) Insulitis and type I diabetes – lessons from the NOD mouse Academia pages. New York London and 12, 152
- mouse. Academic press. New York London, pp 143-153
 41. Maruyama T, Takei I, Taniyama M, Katoaka K, Matsuki S (1984)
 Immunological aspect of non-obese diabetic mice: immune islet
 cell killing mechanism and cell-mediated immunity. Diabetologia
 27: 121-125
- Koike T, Itoh Y, Ishii T, Ito I, Takabayashi K, Maruyama N, Tomioka H, Yoshida S (1987) Preventive effect of monoclonal anti-L3T4 antibody on development of diabetes in NOD mice. Diabetes 36: 539-541
- Wang Y, Hao L, Gill RG, Lafferty KJ (1987) Autoimmune diabetes in NOD mouse is L3T4 T-lymphocyte dependent. Diabetes 36: 535-538
- Charlton B, Bacelj A, Mandel TE (1988) Administration of silica particles or anti-Lyt2 antibody prevents beta-cell destruction in NOD mice given cyclophosphamide. Diabetes 37: 930-935
- Harada M, Makino S (1984) Promotion of spontaneous diabetes in non-obese diabetes-prone mice by cyclophosphamide. Diabetologia 27: 604-606
- Bosi E, Todd I, Pujol-Borrell R, Bottazzo GF (1987) Mechanisms of autoimmunity: relevance to the pathogenesis of Type I diabetes. Diabetes Metab Rev 3: 893-928
- Foulis AK, Farquharson MA, Hardman R (1987) Aberrant expression of class II major histocompatibility complex molecules

- by B cells and hyperexpression of class I major histocompatibility complex molecules by insulin containing islets in Type 1 (insulindependent) diabetes mellitus. Diabetologia 30: 333-343
- Hanafusa Ť, Fujino-Kurihara H, Miyazaki A, Yamada K, Nakajima H, Miyagawa J, Kono N, Tarui S (1987) Expression of class II major histocompatibility complex antigens on pancreatic B cells in the NOD mouse. Diabetologia 30: 104-148
- Momburg F, Koch N, Moller P, Moldenhauser G, Butcher G, Hammerling GJ (1986) Differential expression of Ia and Ia-associated invariant chain in mouse tissues after in vivo treatment with IFN-gamma. J Immunol 136: 940-948
- Kida K, Kaino Y, Miyagawa T, Gotoh Y, Matsuda H, Kono T (1986) Effect of cyclosporin on insulitis and ICSA in NOD mic. In: Taruis, Tochino Y, Nonaka K (eds) Insulitis and Type 1 diabetes lessons from the NOD mouse. Academic Press, New York London, pp 137-142
- Formby B, Miller N, Garret R, Peterson CM (1987) Effects of low-dose cyclosporine prophylaxis in nonobese diabetic mice. J Pharm Exp Ther 241: 1106–1111
- 52 Yamada K, Nonaka K, Hanafusa T, Miyazaki A, Toyoshima H, Tarui S (1982) Preventive and therapeutic effects of large-dose nicotinamide injections on diabetes associated with insultifs – an observation in non obese diabetic (NOD) mice. Diabetes 31: 749-753
- 53. Nakajima H, Yamada K, Hanafusa T, Fujino-Kurihara H, Miyagawa J, Miyazaki A, Saito R, Minami Y, Kono N, Nonaka Tochino Y, Tarui S (1986) Elevated antibody-dependent cell-mediated cytotoxicity and its inhibition by nicotinamide in the diabetic NOD mouse. Immunol Lett 12: 91-94
- Nomikos IN, Prows SJ, Carotenuto P, Lafferty KJ (1986) Combined treatment with nicotinamide and desferrioxamine prevents islet allograft destruction in NOD mice. Diabetes 35: 1302–1304
- Vague P, Vialettes B, Lassman-Vague V, Vallo J (1987) Nicotinamide may extend remission phase in insulin-dependent diabetes. Lancet 1: 619–620
- Mendola G, Casamitjana R, Gomis R (1989) Effect of nicotinamide therapy upon B-cell function in newly diagnosed Type 1 (insulin-dependent) diabetic patients. Diabetologia 32: 160-162
- Pozzilli P, Visalli N, Ghirlanda G, Manna R, Andreani D (1989) Nicotinamide increases C peptide secretion in patients with newly-diagnosed Type 1 (insulin-dependent) diabetes. Diabetic Med 6: 568-572

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